

INTENDED USE

This mouse plasminogen activator inhibitor type 1 (PAI-1) total assay is intended for the quantitative determination of total PAI-1 in mouse plasma in citrate, EDTA or heparin, serum, cell culture media, tissue lysates and other biological fluids. **For research use only.**

BACKGROUND

PAI-1 is a central regulator of the blood fibrinolytic system [1]. Clinical studies have indicated that increased PAI-1 levels increase the risk for thrombosis, whereas decreased levels may cause recurrent bleeding [2].

ASSAY PRINCIPLE

Mouse PAI-1 will bind to the monoclonal capture antibody coated on the microtiter plate. Free, latent, and complexed PAI-1 will bind to the plate. After appropriate washing steps, polyclonal anti-mouse PAI-1 primary antibody binds to the captured protein. Excess primary antibody is washed away and bound antibody is reacted with peroxidase conjugated secondary antibody. Following an additional washing step, TMB substrate is used for color development at 450nm. A standard calibration curve is prepared along with the samples to be measured using dilutions of mouse PAI-1. Color development is proportional to the concentration of PAI-1 in the samples.

REAGENTS PROVIDED

- **96-well antibody coated microtiter strip plate** (removable wells 8x12) containing anti-mouse PAI-1 antibody, blocked and dried.
- **10X Wash buffer:** 1 bottle of 50ml
- **Mouse PAI-1 standard:** 1 vial lyophilized standard
- **Anti-mouse PAI-1 primary antibody:** 1 vial lyophilized polyclonal antibody
- **Anti-rabbit horseradish peroxidase secondary antibody:** 1 vial concentrated HRP labeled antibody
- **TMB substrate solution:** 1 bottle of 10ml solution

STORAGE AND STABILITY

Store all kit components at 4°C upon arrival. Return any unused microplate strips to the plate pouch with desiccant. Reconstituted standards and primary may be stored at -80°C for later use. Do not freeze-thaw the standard and primary antibody more than once. Store all other unused kit components at 4°C. This kit should not be used beyond the expiration date.

OTHER REAGENTS AND SUPPLIES REQUIRED

- Microtiter plate shaker capable of 300 rpm uniform horizontally circular movement
- Manifold dispenser/aspirator or automated microplate washer
- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes and Pipette tips
- Deionized or distilled water
- Polypropylene tubes for dilution of standard
- Paper towels or laboratory wipes
- 1N H₂SO₄ or 1N HCl
- Bovine Serum Albumin Fraction V (BSA)
- Tris(hydroxymethyl)aminomethane (Tris)
- Sodium Chloride (NaCl)

PRECAUTIONS

- FOR LABORATORY RESEARCH USE ONLY. NOT FOR DIAGNOSTIC USE.
- Do not mix any reagents or components of this kit with any reagents or components of any other kit. This kit is designed to work properly as provided.
- Always pour peroxidase substrate out of the bottle into a clean test tube. Do not pipette out of the bottle as contamination could result.
- Keep plate covered except when adding reagents, washing, or reading.
- DO NOT pipette reagents by mouth and avoid contact of reagents and specimens with skin.
- DO NOT smoke, drink, or eat in areas where specimens or reagents are being handled.

PREPARATION OF REAGENTS

- TBS buffer:** 0.1M Tris, 0.15M NaCl, pH 7.4
- Blocking buffer (BB):** 3% BSA (w/v) in TBS
- 1X Wash buffer:** Dilute 50ml of 10X wash buffer concentrate with 450ml of deionized water

SAMPLE COLLECTION

Collect plasma using citrate, EDTA or heparin as an anticoagulant. Immediately after collection of blood, samples must be centrifuged at 3000xg for 15 minutes. It is important to ensure a platelet free preparation since platelets can release PAI-1 [3].

ASSAY PROCEDURE

Perform assay at room temperature. Vigorously shake plate (300rpm) at each step of the assay.

Preparation of Standard

Reconstitute standard by adding 5ml of blocking buffer directly to the vial and agitate gently to completely dissolve contents. This will result in a 50ng/ml standard solution.

Dilution table for preparation of mouse PAI-1 standard:

PAI-1 concentration (ng/ml)	Dilutions
50	100µl from standard vial
20	600µl (BB) + 400µl (50ng/ml)
10	500µl (BB) + 500µl (20ng/ml)
5	500µl (BB) + 500µl (10ng/ml)
2.5	500µl (BB) + 500µl (5ng/ml)
1	600µl (BB) + 400µl (2.5ng/ml)
0.5	500µl (BB) + 500µl (1ng/ml)
0.2	600µl (BB) + 400µl (0.5ng/ml)
0.1	500µl (BB) + 500µl (0.2ng/ml)
0.05	500µl (BB) + 500µl (0.1ng/ml)
0	500µl (BB) Zero point to determine background

NOTE: DILUTIONS FOR THE STANDARD CURVE AND ZERO STANDARD MUST BE MADE AND APPLIED TO THE PLATE IMMEDIATELY.

Standard and Unknown Addition

Remove microtiter plate from bag and add 100µl PAI-1 standards (in duplicate) and unknowns to wells. Carefully record position of standards and unknowns. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

NOTE: The assay measures PAI-1 antigen in the 0.05-50 ng/ml range. If the unknown is thought to have high PAI-1 levels, dilutions may be made in plasma devoid of PAI-1 (cat# MPLA-SC-PAI) or in blocking buffer. A 1:5 dilution for normal plasma is suggested for best results.

Primary Antibody Addition

Reconstitute primary antibody by adding 10ml of blocking buffer directly to the vial and agitate gently to completely dissolve contents. Add 100µl to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

Secondary Antibody Addition

Briefly centrifuge vial before opening. Dilute 1µl of conjugated secondary antibody in 10ml of blocking buffer and add 100µl to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

Substrate Incubation

Add 100µl TMB substrate to all wells and shake plate for 1-5 minutes. Substrate will change from colorless to different strengths of blue. Quench reaction by adding 50µl of 1N H₂SO₄ or HCl stop solution to all wells when samples are visually in the same range as the standards. Add stop solution to wells in the same order as substrate upon which color will change from blue to yellow. Mix thoroughly by gently shaking the plate.

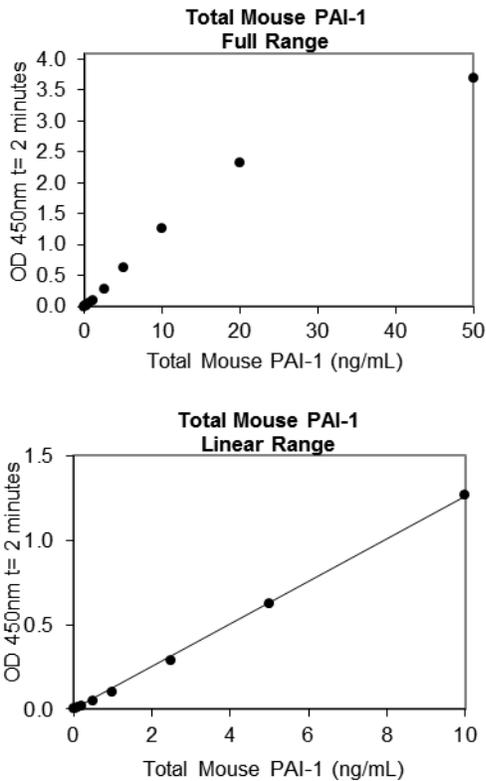
Measurement

Set the absorbance at 450nm in a microtiter plate spectrophotometer. Measure the absorbance in all wells at 450nm. Subtract zero point from all standards and unknowns to determine corrected absorbance (A₄₅₀).

Calculation of Results

Plot A₄₅₀ against the amount of PAI-1 in the standards. Fit a straight line through the linear points of the standard curve using a linear fit procedure if unknowns appear on the linear portion of the standard curve. Alternatively, create a standard curve by analyzing the data using a software program capable of generating a four parameter logistic (4PL) curve fit. The amount of PAI-1 in the unknowns can be determined from this curve. If samples have been diluted, the calculated concentration must be multiplied by the dilution factor.

A typical standard curve (EXAMPLE ONLY):



EXPECTED VALUES

In house testing of pooled normal mouse plasma results:

Strain	Active PAI-1	Total PAI-1
CD-1	1.03 ng/ml	17.55 ng/ml

The concentration level of PAI-1 antigen in mouse plasma was found to be 1.9 ± 0.6 ng/ml [4]. Abnormalities in PAI-1 levels have been reported in the following conditions:

- Endotoxemia: Endotoxin induces a large increase in PAI-1 levels (80-fold) [4].
- Hyperglycemia, hyperinsulinemia, and insulin resistance: Elevated PAI-1 levels in obese and diabetic mice contribute to these metabolic disorders [5,6].
- Vascular thrombosis: Increased PAI-1 levels may contribute to venous thrombosis [1].
- Myocardial Infarction: Increased PAI-1 levels may contribute to myocardial infarction [1].

PERFORMANCE CHARACTERISTICS

Sensitivity: The minimum detectable dose (MDD) was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates (range OD₄₅₀: 0.046-0.053) and calculating the corresponding concentration. The MDD was 0.031 ng/ml.

Intra-assay Precision: Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Sample	1	2	3
n	20	20	20
Mean (ng/ml)	0.288	1.64	20.8
Standard Deviation	0.023	0.081	1.86
CV (%)	7.90	4.92	8.95

Inter-assay Precision: Three samples of known concentration were tested in ten independent assays to assess inter-assay precision.

Sample	1	2	3
n	10	10	10
Mean (ng/ml)	0.307	1.46	16.6
Standard Deviation	0.040	0.155	2.41
CV (%)	12.9	10.6	14.6

Recovery: The recovery of antigen spiked to levels throughout the range of the assay in PAI-1 depleted plasma was evaluated.

Sample	1	2	3	4
n	4	4	4	4
Mean (ng/ml)	0.442	1.59	3.57	15.1
Average % Recovery	109	105	90	99
Range	104-112%	102-107%	89-92%	97-103%

Linearity: To assess the linearity of the assay, pooled citrated mouse plasma samples containing high concentrations of antigen were serially diluted to produce samples with values within the dynamic range of the assay.

Sample	1:2	1:4	1:8	1:16
n	4	4	4	4
Average % of Expected	102	86.9	97.0	105
Range	96.2-107%	85.3-87.5%	94.9-100%	102-107%

Specificity: This assay recognizes natural and recombinant mouse PAI-1. Pooled normal plasma from rat, human, pig, dog, and sheep were assayed, and no significant cross-reactivity was observed. Pooled normal plasma from rabbit resulted in significant background color development.

DISCLAIMER

This information is believed to be correct but does not claim to be all-inclusive and shall be used only as a guide. The supplier of this kit shall not be held liable for any damage resulting from handling of or contact with the above product.

Sample Values: Samples were evaluated for the presence of the antigen at varying dilutions.

REFERENCES

1. Eitzman DT, *et al.*: Blood. 2000, 95(2): 577-580.
2. Kawasaki T, *et al.*: Blood. 2000, 96(1): 153-160.
3. Declerck PJ, *et al.*: Blood. 1988, 71(1): 220-225.
4. Declerck PJ, *et al.*: Thromb Haemostas. 1995, 74(5): 1305-9.
5. Schafer K, *et al.*: FASEB. 2001, 15: 1840-2.
6. Samad F, *et al.*: PNAS. 1996, 96(12): 6902-7.

Sample Type	Dilution	Mean (ng/mL)
Citrate Plasma	1:5	32
	1:10	31
EDTA Plasma	1:5	13
	1:10	13
Heparin Plasma	1:5	19
	1:10	20

Example of ELISA Plate Layout

96 Well Plate: 22 Standard wells, 74 Sample wells

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0.05 ng/ml	0.1 ng/ml	0.2 ng/ml	0.5 ng/ml	1 ng/ml	2.5 ng/ml	5 ng/ml	10 ng/ml	20 ng/ml	50 ng/ml	
B	0	0.05 ng/ml	0.1 ng/ml	0.2 ng/ml	0.5 ng/ml	1 ng/ml	2.5 ng/ml	5 ng/ml	10 ng/ml	20 ng/ml	50 ng/ml	
C												
D												
E												
F												
G												
H												