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Mouse Prekallikrein Total Antigen ELISA Kit

Catalog # MPKKT-TOT

Strip well format. Reagents for up to 96 tests.

Rev: March 2018

SAMPLE INSERT
Refer to kit box for lot specific instructions

INTENDED USE

This mouse prekallikrein total antigen assay is intended for the quantitative determination of total prekallikrein antigen in mouse plasma. **For research use only.**

BACKGROUND

Prekallikrein is the glycosylated single chain zymogen precursor of the plasma serine protease kallikrein. Plasma prekallikrein circulates with kininogen and is activated by Factor XIIa in the intrinsic coagulation pathway. Kallikrein activates plasminogen in fibrinolysis and cleaves kininogen in the bradykinin system of vasodilation. Prekallikrein deficiency is rare and causes increased activated partial thromboplastin time [1]. Elevated plasma prekallikrein is associated with diabetes [2] and cardiovascular disease [3].

ASSAY PRINCIPLE

Mouse prekallikrein will bind to the monoclonal capture antibody coated on the microtiter plate. Prekallikrein and kallikrein will react with the antibody on the plate. After appropriate washing steps, anti-mouse prekallikrein primary antibody binds to the captured protein. Excess primary antibody is washed away and bound antibody is reacted with horseradish peroxidase conjugated streptavidin. Following an additional washing step, TMB substrate is used for color development at 450nm. A standard calibration curve is prepared along with the samples to be measured using dilutions of mouse prekallikrein. Color development is proportional to the concentration of total prekallikrein in the samples.

REAGENTS PROVIDED

- **96-well antibody coated microtiter strip plate** (removable wells 8x12) containing anti-mouse prekallikrein antibody, blocked and dried.
- **10X Wash buffer:** 1 bottle of 50ml
- **Mouse prekallikrein standard:** 1 vial lyophilized standard
- **Anti-mouse prekallikrein primary antibody:** 1 vial lyophilized monoclonal antibody
- **Horseradish peroxidase-conjugated streptavidin:** 1 vial concentrated HRP labeled streptavidin
- **TMB substrate solution:** 1 bottle of 10ml solution

STORAGE AND STABILITY

Store all kit components at 4°C upon arrival. Return any unused microplate strips to the plate pouch with desiccant. Reconstituted standards and primary may be stored at -80°C for later use. Do not freeze-thaw the standard and primary antibody more than once. Store all other unused kit components at 4°C. This kit should not be used beyond the expiration date.

OTHER REAGENTS AND SUPPLIES REQUIRED

- Microtiter plate shaker capable of 300 rpm uniform horizontally circular movement
- Manifold dispenser/aspirator or automated microplate washer
- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes and Pipette tips
- Deionized or distilled water
- Polypropylene tubes for dilution of standard
- Paper towels or laboratory wipes
- 1N H_2SO_4 or 1N HCl
- Bovine Serum Albumin Fraction V (BSA)
- Tris(hydroxymethyl)aminomethane (Tris)
- Sodium Chloride (NaCl)

PRECAUTIONS

- FOR LABORATORY RESEARCH USE ONLY. NOT FOR DIAGNOSTIC USE.
- Do not mix any reagents or components of this kit with any reagents or components of any other kit. This kit is designed to work properly as provided.
- Always pour peroxidase substrate out of the bottle into a clean test tube. Do not pipette out of the bottle as contamination could result.
- Keep plate covered except when adding reagents, washing, or reading.
- DO NOT pipette reagents by mouth and avoid contact of reagents and specimens with skin.
- DO NOT smoke, drink, or eat in areas where specimens or reagents are being handled.

PREPARATION OF REAGENTS

- **TBS buffer:** 0.1M Tris, 0.15M NaCl, pH 7.4
- **Blocking buffer (BB):** 3% BSA (w/v) in TBS
- **1X Wash buffer:** Dilute 50ml of 10X wash buffer concentrate with 450ml of deionized water

SAMPLE COLLECTION

Collect plasma using EDTA or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000xg within 30 minutes of collection. Assay immediately or aliquot and store at ≤ -20°C. Avoid repeated freeze-thaw cycles.

ASSAY PROCEDURE

Perform assay at room temperature. Vigorously shake plate (300rpm) at each step of the assay.

Preparation of Standard

Reconstitute standard by adding 1ml of blocking buffer directly to the vial and agitate gently to completely dissolve contents. This will result in a 100ng/ml standard solution.

Dilution table for preparation of mouse prekallikrein standard:

Prekallikrein concentration (ng/ml)	Dilutions
100	Straight from vial
50	500µl (BB) + 500µl (from vial)
25	500µl (BB) + 500µl (50ng/ml)
10	600µl (BB) + 400µl (25ng /ml)
5	500µl (BB) + 500µl (10ng /ml)
2	600µl (BB) + 400µl (5ng /ml)
1	500µl (BB) + 500µl (2ng /ml)
0.5	500µl (BB) + 500µl (1ng /ml)
0.2	600µl (BB) + 400µl (0.5ng /ml)
0.1	500µl (BB) + 500µl (0.2ng /ml)
0	500µl (BB) Zero point to determine background

NOTE: DILUTIONS FOR THE STANDARD CURVE AND ZERO STANDARD MUST BE MADE AND APPLIED TO THE PLATE IMMEDIATELY.

Standard and Unknown Addition

Remove microtiter plate from bag and add 100µl prekallikrein standards (in duplicate) and unknowns to wells. Carefully record position of standards and unknowns. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

NOTE: The assay measures mouse prekallikrein antigen in the 0.1-100ng/ml range. If the unknown is thought to have high prekallikrein levels, dilutions may be made in blocking buffer. A 1:10,000-1:20,000 dilution for normal mouse plasma is suggested for best results.

Primary Antibody Addition

Reconstitute primary antibody by adding 10ml of blocking buffer directly to the vial and agitate gently to completely dissolve contents. Add 100µl to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

Streptavidin-HRP Addition

Briefly centrifuge vial before opening. Dilute 2.5µl of HRP conjugated streptavidin into 2.5ml blocking buffer to generate a 1:1,000 dilution. Add 0.4ml of 1:1,000 dilution to 9.6ml of blocking buffer to generate a 1:25,000 dilution. Add 100µl of the 1:25,000 dilution to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

Substrate Incubation

Add 100µl TMB substrate to all wells and shake plate for 2-10 minutes. Substrate will change from colorless to different strengths of blue. Quench reaction by adding 50µl of 1N H_2SO_4 or HCl stop solution to all wells when samples are visually in the same range as the standards. Add stop solution to wells in the same order as substrate upon which color will change from blue to yellow. Mix thoroughly by gently shaking the plate.

Measurement

Set the absorbance at 450nm in a microtiter plate spectrophotometer. Measure the absorbance in all wells at 450nm. Subtract zero point from all standards and unknowns to determine corrected absorbance (A_{450}).

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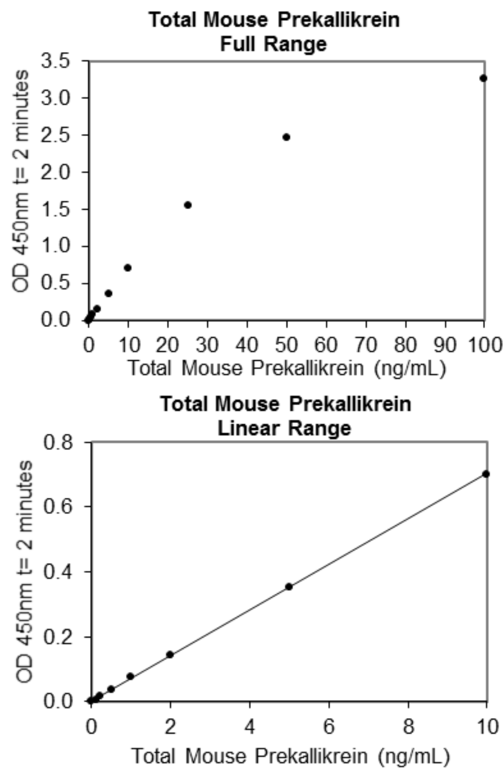
Calculation of Results

Plot A_{450} against the amount of prekallikrein in the standards. Fit a straight line through the linear points of the standard curve using a linear fit procedure if unknowns appear on the linear portion of the standard curve. Alternatively, create a standard curve by analyzing the data using a software program capable of generating a four parameter logistic (4PL) curve fit. The amount of prekallikrein in the unknowns can be determined from this curve. If samples have been diluted, the calculated concentration must be multiplied by the dilution factor.

A typical standard curve (EXAMPLE ONLY):



EXPECTED VALUES

Prekallikrein is present in normal human plasma at concentrations of 15µg/ml [4] to 55µg/ml [5] as determined by clotting assay. Prekallikrein antigen measured by ELISA has not been reported. Normal values of prekallikrein in mouse plasma have not been conclusively determined but are believed to be similar to human plasma based on in-house testing.

PERFORMANCE CHARACTERISTICS

Sensitivity: The minimum detectable dose (MDD) was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates (range OD450: 0.045-0.049) and calculating the corresponding concentration. The MDD was 0.044ng/ml.

Intra-assay Precision: Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Sample	1	2	3
n	20	20	20
Mean (ng/ml)	0.48	1.29	6.25
Standard Deviation	0.035	0.063	0.25
CV (%)	7.36	4.87	3.99

Inter-assay Precision: These studies are currently in progress. Please contact us for more information.

Recovery: These studies are currently in progress. Please contact us for more information.

Linearity: To assess the linearity of the assay, samples containing high concentrations of antigen were serially diluted to produce samples with values within the dynamic range of the assay.

Sample	1:2	1:4	1:8	1:16
n	4	4	4	4
Average % of Expected	99	100	101	100
Range	98-100%	99-101%	98-103%	95-104%

Specificity: This assay recognizes total mouse prekallikrein. Pooled normal plasma from rat, rabbit, dog, pig, sheep, horse, human, cyno monkey and rhesus monkey was assayed and no significant cross-reactivity was observed.

Sample Values: Samples were evaluated for the presence of the antigen at varying dilutions.

Sample Type	Dilution	Mean (µg/mL)
Citrate Plasma	1:8,000	59.7
	1:16,000	59.6
	1:32,000	61.8
	1:64,000	62.8
	1:128,000	62.2

DISCLAIMER

This information is believed to be correct but does not claim to be all-inclusive and shall be used only as a guide. The supplier of this kit shall not be held liable for any damage resulting from handling of or contact with the above product.

REFERENCES

1. Sollo DG and Saleem A: Ann Clin Lab Sci. 1985, 15:279-285.
2. Jaffa AA *et al*.: Diabetes. 2003, 52:1215-1221.
3. Mackenzie JA *et al*.: Appl Physiol Nutr Metab. 2010, 35:518-525.
4. Heimark RL and Davie EW: Methods Enzymol. 1981, 80(Pt C):157-172.
5. Bouma AA *et al*.: Biochemistry. 1980, 19:1151-1160.

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Example of ELISA Plate Layout
96 Well Plate: 22 Standard wells, 74 Sample wells

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0.1 ng/ml	0.2 ng/ml	0.5 ng/ml	1 ng/ml	2 ng/ml	5 ng/ml	10 ng/ml	25 ng/ml	50 ng/ml	100 ng/ml	
B	0	0.1 ng/ml	0.2 ng/ml	0.5 ng/ml	1 ng/ml	2 ng/ml	5 ng/ml	10 ng/ml	25 ng/ml	50 ng/ml	100 ng/ml	
C												
D												
E												
F												
G												
H												